

Original Article

α -Tocopheryl succinate induces apoptosis in erbB2-expressing breast cancer cell via NF- κ B pathway

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Aim: To study the molecular mechanisms underlying α -tocopheryl succinate (α -TOS)-induced apoptosis in erbB2-positive breast cancer cells and to determine whether α -TOS and the human recombinant TNF-related apoptosis-inducing ligand (hrTRAIL) act synergically to induce cell death of erbB2-expressing breast cancer cells.

Methods: The annexin V binding method was used to measure apoptosis induced by α -TOS and/or hrTRAIL. RT-PCR and Western blotting were performed to detect gene and protein expression. A colorimetric assay was performed to detect caspase activity. The TransAM™ NF- κ B p65 kit was used to assess NF- κ B activation.

Results: α -TOS (100 μ mol/L) significantly inhibited NF- κ B nuclear translocation in erbB2-expressing breast cancer cells; this inhibition is expected to result in the inactivation of NF- κ B. α -TOS (50 and 100 μ mol/L) inhibited the expression of Flice-like inhibitory protein (FLIP) and cellular inhibitor of apoptosis protein 1 (c-IAP1) in erbB2-positive cells. α -TOS (100 μ mol/L) inhibited Akt activation and augmented the activity of caspase 3 and caspase 8 in breast cancer cells expressing erbB2. α -TOS (50 μ mol/L) and hrTRAIL (30 mg/mL) acted synergically to induce apoptosis in breast cancer cells. α -TOS also decreased the hrTRAIL-induced transient activation of NF- κ B.

Conclusion: Our results suggest that α -TOS mediates the apoptosis of erbB2-positive breast cancer cells and acts synergically with hrTRAIL via the NF- κ B pathway.

Keywords: erbB2; α -TOS; breast cancer; NF- κ B; apoptosis; hrTRAIL

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Introduction

Breast cancer cells that express erbB2 are resistant to many drugs^[1, 2]. The major complication associated with erbB2 expression is linked to auto-activation of downstream signaling pathway(s), such as phosphatidylinositol 3-kinase (PI3K), which leads to the activation of Akt^[3], a serine/threonine kinase that promotes cellular survival^[4]. Akt causes activation of the transcription factor nuclear factor- κ B (NF- κ B)^[5, 6]. In most non-transformed cells, NF- κ B complexes (heterotrimers usually composed of the p50 and p65 subunits bound to an inhibitor subunit, I κ B) are largely cytoplasmic. Activation of NF- κ B causes it to translocate into the nucleus and bind to the promoter regions of specific pro-survival genes, such as those coding for IAPs^[7] and the caspase 8 inhibitor, FLIP^[8]. There-

fore, agents that inhibit NF- κ B activation in erbB2-expressing cancer cells are of clinical interest.

Vitamin E (VE) analogs, a novel class of VE compounds with strong pro-apoptotic activity epitomized by α -TOS, have been shown to trigger apoptosis in a variety of cancer cells^[9]. Recently, we found that α -TOS induced apoptosis in breast cancer cells expressing erbB2^[10]. Using pre-clinical models, it has been shown that hrTRAIL is a potent anticancer agent^[11]. However, some tumor cells possess intrinsic resistance to hrTRAIL^[12], one of the reasons for this is that hrTRAIL can induce transient NF- κ B activation in some cell types^[13]. A cooperative apoptotic effect of α -TOS and hrTRAIL has been observed in human malignant mesothelioma cells^[11].

To determine the molecular mechanisms by which α -TOS induces apoptosis in erbB2-positive breast cancer cells and synergizes with hrTRAIL, we measured the activation of Akt and NF- κ B, the nuclear translocation of NF- κ B p65, the caspase activity and the expression of caspase inhibitors. We found that the NF- κ B pathway plays a critical role in α -TOS-induced

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breast cancer cell apoptosis and its synergy with hrTRAIL.

Materials and methods

Cell culture and treatment

Human breast cancer cell lines MCF7 and MDA-MB-453 were routinely cultured in DMEM supplemented with 10% FCS, 100 U/mL penicillin and 100 µg/mL streptomycin at 37 °C in a humidified atmosphere with 5% CO₂. At approximately 50% to 70% confluence, cells were treated with up to 100 µmol/L α-TOS (Sigma Chemical Co, Missouri, USA) and/or up to 30 ng/mL hrTRAIL (Sigma Chemical Co, Missouri, USA)^[14]. After exposure to the drugs for the required period of time, cells were harvested and assessed by different methods (see below).

Western blotting

Whole cell lysates were obtained using a kit (Beijing SBS Genetech Co, Ltd, Beijing, China). Nuclear and cytoplasmic fractions were obtained using the Nuclear and Cytoplasmic Protein Extraction Kit (KeyGEN, Nanjing, China). Protein samples were denatured, resolved in 10% SDS-polyacrylamide gels and transferred onto nitrocellulose membranes. The membranes were blocked and incubated with primary antibodies (Santa Cruz, California, USA) then incubated further with horseradish peroxidase (HRP)-conjugated secondary IgG. Anti-β-actin or anti-histone 1 (Santa Cruz, California, USA) was used as a control for protein loading.

Assessment of markers of apoptosis

Apoptosis was quantified by flow cytometry using the Annexin V method as described elsewhere^[10]. Cells were plated at a seeding density of 10⁵ cells/well in 24-well plates. After overnight incubation, cells were treated with α-TOS and/or hrTRAIL for varying durations. Both floating and attached cells were harvested, washed with PBS, resuspended in 0.2 mL binding buffer (10 mmol/L HEPES, 140 mmol/L NaCl, and 5 mmol/L CaCl₂, pH 7.4), incubated for 30 min at 4 °C with 2 µL annexin V-FITC, supplemented with 200 µL propidium iodide (PI) (50 µg/mL), and analyzed by flow cytometry using channel 1 for annexin V-FITC binding and channel 2 for PI staining. Cell death was consistently quantified as the percentage of cells with high annexin V binding plus PI staining.

RNA interference (RNAi)

Validated, erbB2-specific short interfering RNA (siRNA) oligonucleotides and nonspecific scrambled siRNA were purchased from Ambion (Austin, TX, USA). Briefly, cells were allowed to reach 50% confluence, supplemented with 60 pmol/L siRNA pre-incubated with Oligofectamine (Invitrogen, Carlsbad, CA, USA) and overlaid with OptiMEM. Cells were washed 24 h later with PBS, overlaid with complete DMEM, and cultured for an additional 24 h. Knockdown efficiency was confirmed by Western blot before using the cells for further experiments^[15].

RT-PCR

Total RNA was extracted from cells using Trizol Reagent (Invitrogen, Carlsbad, CA, USA). Reverse transcription was carried out using AMV reverse transcriptase (Invitrogen, Carlsbad, CA, USA) and oligo(dT)18 primer. One to three microliters of the cDNA was used for amplification, and PCR was run for 30 cycles. The specific oligonucleotide primer pairs for certain genes and the expected sizes of the PCR products are as follows: c-IAP1 FW, 5'-GAA TAC TCC CTG TGA TTA ATG GTG CCG TGG-3' and RV, 5'-TCT CTT GCT TGT AAA GAC GTC TGT GTC TTC-3', 230 bp; FLIP FW, 5'-CAT ACT GAG ATG CAA GAA TT-3' and RV, 5'-GCT GAA GTC ATC CAT GAG GT-3', 875 bp; β-actin FW, 5'-TGA CGG GGT CAC CCA CAC TGT GCC-3' and RV, 5'-CTA GAA GCA TTT GCG GTG GAC GAT GGA GGG-3', 662 bp. The conditions used for PCR amplification were as follows: 94 °C for 30 s, 65 °C for 30 s, and 72 °C for 1 min, 30 cycles in a 25-µL reaction.

Assessment of NF-κB activation

Activation of NF-κB was estimated using the TransAM™ NF-κB p65 Kit (Active Motif, Carlsbad, CA, USA) according to the manufacturer's protocol. Briefly, 5×10⁶ cells treated with α-TOS and/or hrTRAIL or TNF were washed with PBS and spun down. The cells were lysed with NP40 in hypotonic buffer and then centrifuged to separate the nuclei. Nuclear proteins were extracted with complete lysis buffer, transferred into wells containing the immobilized NF-κB (p65) consensus sequence and incubated for 1 h at 37 °C. The wells were washed, and the bound p65 protein was detected by HRP-dependent staining following incubation with anti-p65 IgG and HRP-conjugated secondary IgG. The absorbance at 450 nm, which was assessed by microplate reader, reflected the amount of bound p65^[16].

Caspase 3 and caspase 8 activity assays

The activities of caspase 3 and caspase 8 were measured by cleavage of the chromogenic caspase substrates, Ac-DEVD-pNA (acetyl-Asp-Glu-Val-Asp *p*-nitroanilide) and Ac-IETD-pNA (acetyl-Ile-Glu-Thr-Asp *p*-nitroanilide), respectively^[17]. Approximately 30 µg of total protein was added to reaction buffer containing Ac-DEVD-pNA (2 mmol/L) or Ac-IETD-pNA (2 mmol/L) and incubated for 2 h at 37 °C. The absorbance of yellow pNA cleaved from its corresponding precursor was measured using a spectrometer at 405 nm. The specific caspase activity was normalized by total protein of the cell lysates and expressed as the fold increase of the baseline caspase activity in control cells cultured in DMEM with 10% FBS.

Immunocytochemistry

Cells were placed on sterile coverslips in 12-well plates overnight and then fixed with 100% methanol for 3 min, 1% H₂O₂-methanol for 5 min, 95% methanol for 3 min and 50% methanol for 3 min. The cells were then exposed to anti-p65 IgG (Santa Cruz, California, CA, USA), followed by a biotinylated

secondary antibody. After washing with PBS, the cells were incubated with streptavidin-HRP. Finally, protein expression was detected using DAB solution and observed by microscopy.

Statistical analysis

Data were obtained from at least three different experiments, and the results are presented as the mean \pm SD. Significance between data points was assessed using the Student's *t*-test, and the data were considered significantly different when $P < 0.05$.

Results

α -TOS inhibited NF- κ B nuclear translocation in breast cancer cells expressing erbB2

We determined that both MDA-MB-453 and MCF7 cell lines express erbB2 and phosphorylated Akt (pAkt) and similar levels of Akt (Figure 1A). This result is consistent with previous data showing that the expression of erbB2 leads to activated Akt. Akt activation leads to the activation of NF- κ B, which controls the expression of a variety of antiapoptotic genes. Immunocytochemistry for the p65 protein (a subunit of NF- κ B) showed much weaker nuclear staining in MDA-MB-453 (Figure 1B) and MCF7 (data not shown) cells after treatment with 100 μ mol/L α -TOS than in untreated cells. Western blotting showed that p65 protein decreased in the nuclear fraction and increased in the cytoplasmic fraction after treatment with α -TOS in both cell lines (Figure 1C). We can conclude that

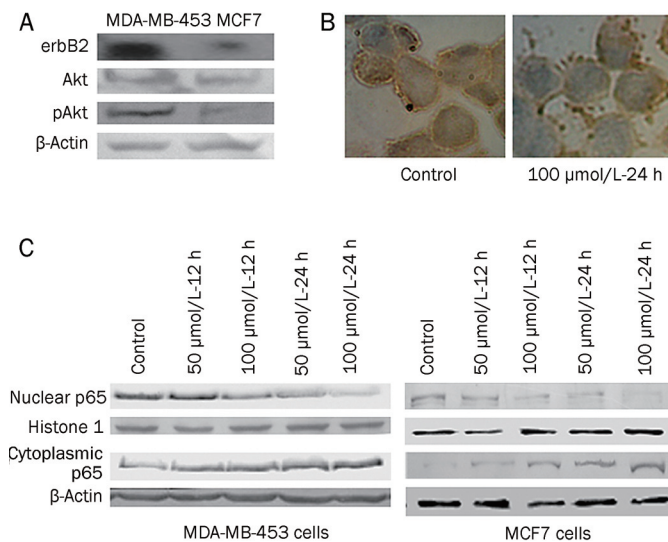


Figure 1. α -TOS inhibited p65 nuclear translocation in erbB2-expressing breast cancer cells. (A) MDA-MB-453 and MCF7 cells were lysed and the whole cell lysate was probed by immunoblotting for the levels of erbB2, Akt and p-Akt. (B) MDA-MB-453 cells were treated with α -TOS for 24 h. Cells were then fixed for p65 immunostaining. (C) Both cell lines were treated with α -TOS as indicated. As shown, nuclear and cytoplasmic fractions (60 μ g protein per sample) were resolved on a 10% polyacrylamide gel and probed by immunoblotting for the expression of p65. The images are representative of three independent experiments.

α -TOS inhibited the translocation of p65 from the cytoplasm to the nucleus in breast cancer cells expressing erbB2.

α -TOS inhibited Akt activation

Because the activation of Akt correlates with its phosphorylation at the Thr³⁰⁸ and Ser⁴⁷³ residues, we examined the amount of Akt and pAkt (Ser⁴⁷³) after treatment with α -TOS. Figure 2 shows that α -TOS treatment decreased the level of pAkt in MCF7 and MDA-MB-453 cells. The level of total Akt was not altered in either cell line after treatment with α -TOS.

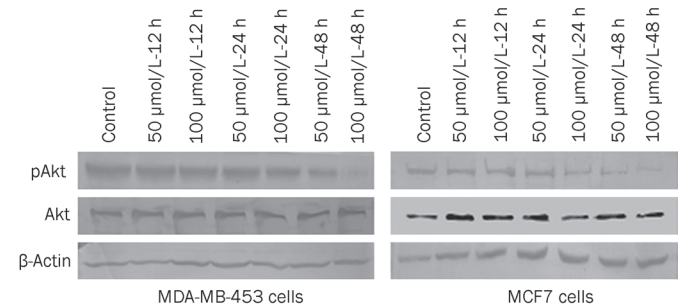


Figure 2. α -TOS inhibited Akt activation in erbB2-expressing breast cancer cells. MCF7 and MDA-MB-453 cells were treated with α -TOS for varying durations. Total cellular proteins were extracted and used in Western blotting for Akt, p-Akt and β -actin.

α -TOS suppressed c-IAP1 and FLIP expression in erbB2-positive breast cancer cells

NF- κ B controls the expression of pro-survival genes such as members of the IAP family^[7] and the caspase 8 inhibitor, FLIP^[8]. This characteristic prompted us to investigate the effects of α -TOS on IAP and FLIP expression. We detected the expression of c-IAP1, c-IAP2 (both members of IAP family) and FLIP after treatment with α -TOS. We determined that α -TOS suppressed c-IAP1 and FLIP expression (Figure 3) but had no effect on c-IAP2 expression (data not shown). Figure 3A shows that α -TOS reduced the amount of c-IAP1 mRNA in MDA-MB-453 and MCF7 cells in a dose- and time-dependent manner. Figure 3B shows that α -TOS reduced c-IAP1 protein expression in both cell lines. A decrease in the amount of FLIP mRNA was also found in α -TOS-treated MDA-MB-453 and MCF7 cells (Figure 3C). Figure 3D shows that α -TOS reduced FLIP protein expression in both cell lines.

α -TOS augmented the activity of caspases

IAPs inhibit distinct members of the caspase family of cysteine proteases. Caspases facilitate apoptosis by cleaving a growing number of cellular targets^[18]. FLIP is an inhibitor of caspase 8^[19]. We found that α -TOS suppressed c-IAP1 and FLIP expression in breast cancer cells expressing erbB2 (Figure 3), which led us to measure activation of caspase 3 and caspase 8. We found that MCF7 cells do not express caspase 3, which is consistent with results reported by Mathiasen^[20]. Figure 4A shows that α -TOS induced an increase in caspase 8 activity in

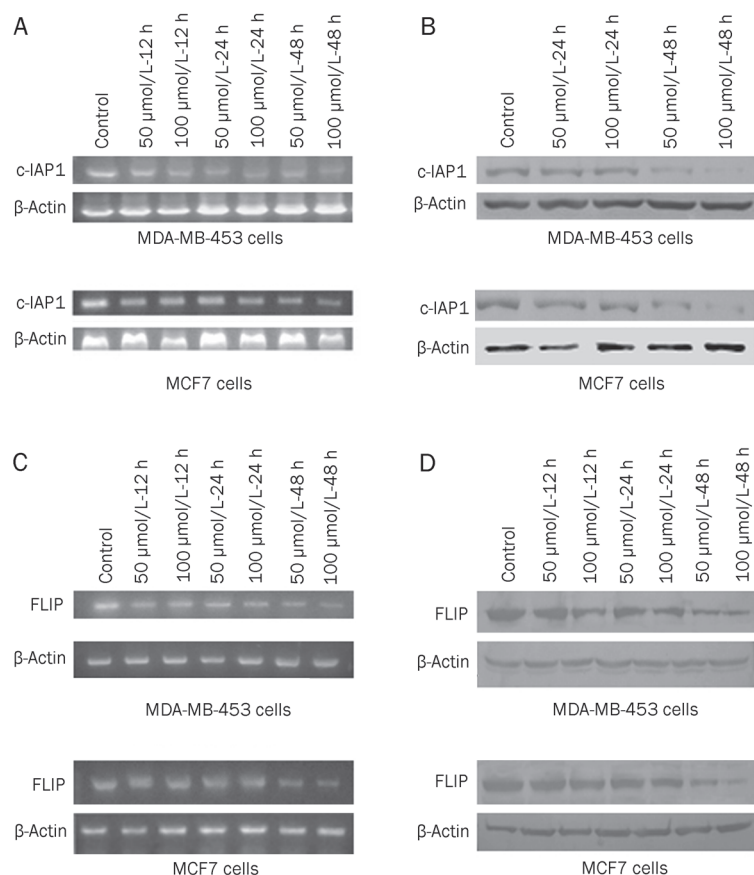


Figure 3. α -TOS inhibited c-IAP1 and FLIP expression in erbB2-expressing breast cancer cells. MDA-MB-453 and MCF7 cells (as shown) were exposed to α -TOS for the durations and at the concentrations shown. They were assessed for the expression of c-IAP1 mRNA by RT-PCR (A), c-IAP1 protein by Western blotting (B), FLIP mRNA by RT-PCR (C) and FLIP protein by Western blotting (D). The images are representative of at least three independent experiments.

MCF7 cells. Figures 4B and 4C show that α -TOS increased the activity of caspase 3 and caspase 8 in MDA-MB-453 cells.

α -TOS induced apoptosis synergically with hrTRAIL

Although hrTRAIL is a potent anticancer agent, it has been reported that erbB2-expressing breast cancer cells are resistant to hrTRAIL^[21]. Figures 5A and 5B show that hrTRAIL induced very low levels of apoptosis in MDA-MB-453 and MCF7 cells, indicating that both of these erbB2-expressing cell lines are

resistant to hrTRAIL. However, both cell lines are sensitive to α -TOS treatment, which is consistent with results reported by Wang^[10], showing that α -TOS causes efficient apoptosis in breast cancer cells regardless of their erbB2 status. Figures 5A and 5B also demonstrate the synergism (or an additive effect) of α -TOS with hrTRAIL in both cell lines. To determine whether the expression of erbB2 was the major reason that cells had low susceptibility to hrTRAIL, we knocked down the level of erbB2 mRNA using erbB2/siRNA in MDA-MB-453

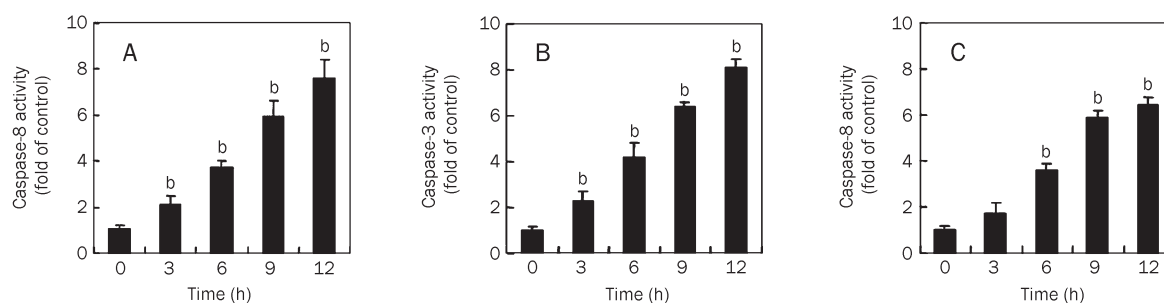


Figure 4. α -TOS augmented caspase activity. (A) MCF7 cells were treated with 100 μ mol/L α -TOS for 0–12 h. Thirty micrograms of total protein were added to reaction buffer containing Ac-IETD-pNA (2 mmol/L) and incubated for 2 h at 37 °C. The absorbance of yellow pNA cleaved from its corresponding precursor was measured using a spectrometer at 405 nm. (B), (C) MDA-MB-453 cells were incubated with 100 μ mol/L α -TOS for 0–12 h. Thirty micrograms of total protein were added to reaction buffer containing Ac-DEVD-pNA (2 mmol/L) or Ac-IETD-pNA (2 mmol/L); the reaction was incubated for 2 h at 37 °C, and the absorbance of yellow pNA cleaved from its corresponding precursor was measured using a spectrometer at 405 nm. The specific caspase activity, normalized to total protein of cell lysates, was then expressed as fold increase compared to baseline caspase activity in the control cells cultured in DMEM with 10% FBS. ^b P <0.05 vs 0 h.

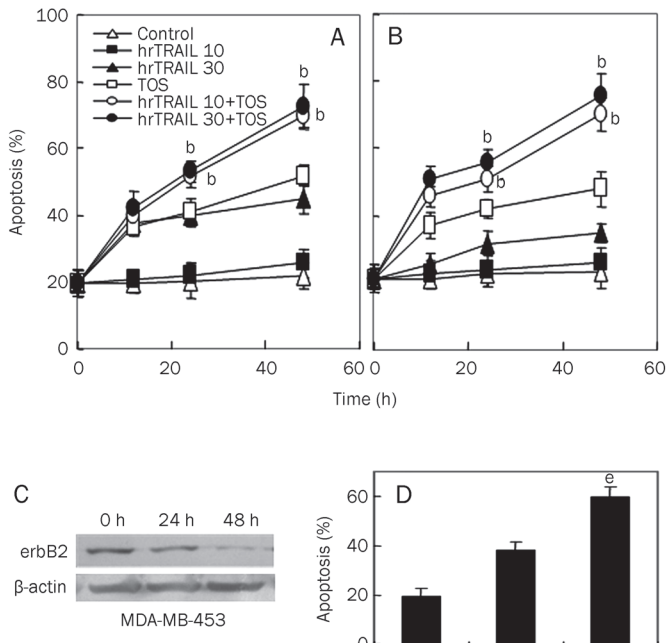


Figure 5. α -TOS acted synergically with hrTRAIL in the induction of apoptosis in erbB2-expressing breast cancer cells. MCF7 (A) and MDA-MB-453 (B) cells were treated with hrTRAIL (10 ng/mL or 30 ng/mL) and/or α -TOS (50 μ mol/L) for the indicated durations. When we detected synergy between hrTRAIL and α -TOS, both reagents were added to the cells at same time. Apoptosis was assessed using the annexin V method and evaluated based on the percentage of annexin V-positive cells scored by FACS analysis. Data are shown as the mean values \pm SD ($n=5-8$). The symbol "b" indicates a significant difference in apoptosis in cells exposed to hrTRAIL+ α -TOS vs cells exposed to hrTRAIL or α -TOS ($^*P<0.05$). (C) MDA-MB-453 cells were evaluated by Western blotting for the expression of erbB2 after treatment with erbB2/siRNA for 24 and 48 h. (D) Induction of apoptosis in control MDA-MB-453 cells (1) and cells pretreated with non-silencing siRNA (2) or erbB2/siRNA (3) and exposed to 30 ng/mL hrTRAIL for 48 h. "e" indicates a significant difference in hrTRAIL-induced apoptosis in cells pretreated with erbB2/siRNA vs cells pretreated with non-silencing siRNA ($^*P<0.05$). The data are derived from three independent experiments and are presented as the mean values \pm SD.

cells that express a higher level of erbB2 than MCF7 cells (Figure 5C). The results, shown in Figure 5D, demonstrate that knocking down the expression of erbB2 sensitized the cells to hrTRAIL-induced apoptosis.

α -TOS decreased the hrTRAIL-induced transient activation of NF- κ B

More direct studies of NF- κ B activation revealed that NF- κ B was activated in MCF7 and MDA-MB-453 cells exposed to hrTRAIL. Figure 6 shows the substantial activation of NF- κ B 30 min after the addition of hrTRAIL to the cells, although this activation was less pronounced than that caused by treatment with the strong NF- κ B activator, TNF- α . This activation was transient and lasted for about 1 h, after which it declined. Pre-treatment with α -TOS abolished the initial NF- κ B activation observed with hrTRAIL alone.

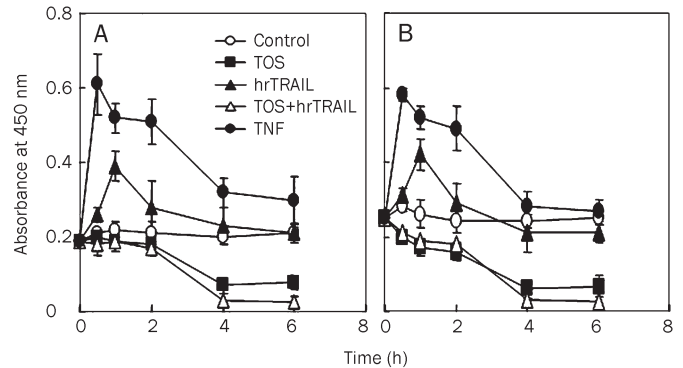


Figure 6. α -TOS decreased the transient activation of NF- κ B caused by hrTRAIL. MCF7 (A) and MDA-MB-453 (B) cells were treated with α -TOS (50 μ mol/L) and/or hrTRAIL (30 ng/mL) or TNF- α (100 U/mL), and washed with PBS. Nuclear proteins were prepared and probed for NF- κ B activation using the TransAMTM NF- κ B p65 kit. The level of NF- κ B activation (p65 binding to its cognate DNA sequence) is expressed as absorbance at 450 nm.

Discussion

In nonmalignant cells, the NF- κ B transcription factor complexes (a heterotrimer usually composed of the p50 and p65 subunits bound to an inhibitor subunit, I κ B) are largely cytoplasmic. Activation of NF- κ B results in the proteasomal degradation of I κ B and nuclear translocation of NF- κ B. Activated NF- κ B then binds to the cognate promoters of genes under its transcriptional control, resulting in their expression.

Activated NF- κ B increases the expression of antiapoptotic genes, such as the caspase-inhibiting IAPs^[7] and FLIP^[8]. Therefore, inhibition of NF- κ B activation may be of great benefit by causing apoptosis in therapy-resistant cancer cells expressing erbB2. Therefore, we hypothesized that α -TOS inhibits NF- κ B activation and promotes apoptosis of erbB2-expressing breast cancer cells. One of the reasons for this hypothesis was the finding that α -TOS causes apoptosis in erbB2-expressing breast cancer cells^[10]. Akazawa and colleagues^[22] also demonstrated that α -tocopheryloxybutyric acid, a compound analog to α -TOS, efficiently causes apoptosis in erbB2-expressing breast cancer cells, although the mechanism by which this occurs was not clarified.

Our studies demonstrate that α -TOS inhibits the NF- κ B pathway based on the following results: (1) α -TOS decreased p65 protein expression in the nuclear fraction of MDA-MB-453 and MCF7 cells (Figure 1C), (2) α -TOS suppressed activation (phosphorylation) of Akt in erbB2-expressing breast cancer cells (Figure 2), (3) α -TOS suppressed c-IAP1 and FLIP expression in erbB2-positive breast cancer cells (Figure 3), and (4) α -TOS increased the activation of caspase 3 and caspase 8 in MDA-MB-453 cells and caspase 8 in MCF7 cells (Figure 4). The above results indicate that α -TOS inhibits the NF- κ B pathway in the breast cancer cells with high (MDA-MB-453) and low (MCF7) expression of erbB2; these findings are consistent with a paper reporting the inhibition of NF- κ B activation by α -TOS in the context of cardiovascular disease^[23].

hrTRAIL has been reported to selectively induce apoptosis *in vitro* and *in vivo*^[24, 25]. A previous study showed that hrTRAIL-activated caspase 8 could generate the truncated Bid protein that triggers the release of cytochrome *c* (Cyt *c*) from mitochondria, leading to assembly of the apoptosome with the ensuing activation of effector caspases. Following the activation of caspase 8, TRAIL can also induce the direct activation of effector caspases^[26]. In both cases, caspase 8 activation is inhibited when the expression of the FLIP protein was upregulated in cancer cells. While the mitochondria are involved in hrTRAIL-induced apoptosis, the IAPs, which are often upregulated in cancer cells, counteract the apoptogenic signals.

Although hrTRAIL is a potent anticancer agent, Matsuzaki *et al*^[12] reported that some tumor cells possess intrinsic or acquired resistance to hrTRAIL. Trauzold *et al*^[27] also demonstrated that the treatment of cells with hrTRAIL resulted in strongly increased distant metastasis of pancreatic tumors *in vivo*. Consistent with these results, we show here that MCF7 and MDA-MB-453 cells were relatively resistant to hrTRAIL-induced apoptosis (Figure 5).

We also document synergy between α -TOS and hrTRAIL in MDA-MB-453 and MCF7 cells (Figure 5). These results are consistent with previous studies in colon cancer cells^[28], in which synergy between hrTRAIL and α -TOS was reported. Similarly, a synergistic effect of hrTRAIL and α -TOS has been observed in mesothelioma cells^[14]. Here, our results indicate that α -TOS decreased the transient activation of NF- κ B caused by hrTRAIL, which may be the molecular mechanism of synergy between hrTRAIL and α -TOS. Dalen and Neuzil^[16] have shown that α -TOS sensitizes cells to TRAIL-induced apoptosis through inhibition of NF- κ B activation. Another publication suggested that the synergy between α -TOS and hrTRAIL occurs through promotion of the mitochondrial apoptotic pathway^[28]. The inhibition of NF- κ B activation by α -TOS has a pro-apoptotic effect that can also be implicated in adjuvant cancer therapy.

Previous studies have shown that α -TOS triggers apoptosis in erbB2-expressing breast cancer cells by signaling via the mitochondrial pathway^[10, 29]. α -TOS induces dissipation of the mitochondrial inner membrane potential and the release of mitochondrial apoptotic proteins such as AIF, Smac/Diablo and Cyt *c*. In many cases, relocation of Cyt *c* leads to formation of the apoptosome, a complex consisting of Cyt *c*, Apaf-1 and procaspase 9, with the ensuing activation of caspase 9 followed by activation of caspase 3^[17, 29].

The data shown in this paper and supported by other reports point to the following mechanism by which α -TOS can overcome the resistance to apoptosis of cancer cells expressing erbB2 (Figure 7). α -TOS suppresses erbB2-dependent activation of NF- κ B, possibly via inhibition of erbB2 auto-activation, as shown elsewhere^[22], which would then lead to inhibition of activation of the central pro-survival factors Akt and/or NF- κ B. Ultimately, this would cause lower levels of expression of the caspase inhibitors FLIP and IAPs, induction of apoptosis by α -TOS and sensitization of cancer cells to other inducers of apoptosis (Figure 7).

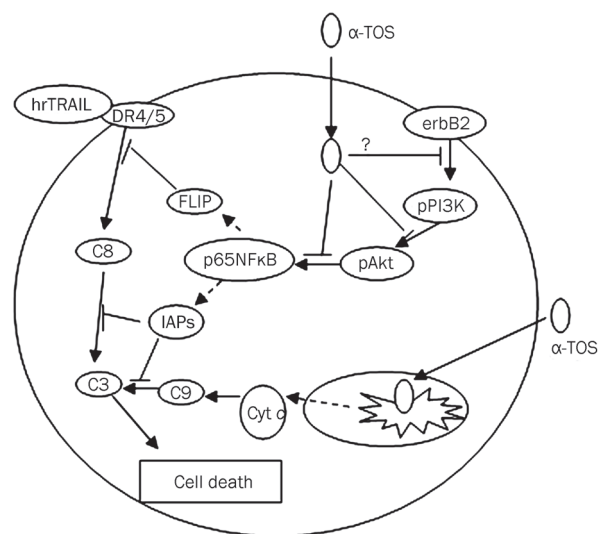


Figure 7. Possible molecular mechanisms by which α -TOS overcomes the resistance of erbB2-expressing cells to apoptosis and sensitizes them to hrTRAIL-induced apoptosis. Expression of erbB2 leads to the activation of the central survival protein, Akt. The kinase then activates a variety of substrates, including NF κ B, which causes increased expression of the caspase inhibitor, FLIP, and IAPs, which inhibit both death receptor-mediated (extrinsic) or mitochondria-dependent (intrinsic) apoptosis. We propose that α -TOS suppresses the activation of NF κ B by a mechanism that may include the inhibition of erbB2, PI3K, or Akt activation. This inhibition results in the decreased expression of FLIP and IAPs and in the initiation of apoptotic signaling by α -TOS and the sensitization of cells to hrTRAIL-mediated apoptosis. C3, C8, and C9: caspase 3, caspase 8, and caspase 9, respectively.

Acknowledgements

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Author contribution

Xiu-fang WANG designed the experiments and wrote the paper; Xiu-fang WANG, Ying XIE, Yuan ZHANG, Hong-gang WANG, Zhan-jun LU, and Xiao-cui DUAN performed the research and analyzed the data.

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